_	218	/		0000 10: 1:-
	210	(gene ADJ silencin\$5) and (animal\$5 or	USPAT;	2002/11/07 11:33
		mammal\$5)	US-PGPUB;	
			EPO; JPO;	
			DERWENT	
-	182	1 , , 3 - ,	USPAT;	2002/09/08 12:53
		mammal\$5)) and repeat\$10	US-PGPUB;	
			EPO; JPO; DERWENT	
_	174	(((gene ADJ silencin\$5) and (animal\$5 or	USPAT;	2002/09/08 12:59
	1/1	mammal\$5)) and repeat\$10) and promoter	US-PGPUB;	2002/03/08 12:33
		manuarys,, and repeaters, and promoter	EPO; JPO;	
			DERWENT	
-	32	((((gene ADJ silencin\$5) and (animal\$5 or	USPAT;	2002/09/08 12:56
		mammal\$5)) and repeat\$10) and promoter)	US-PGPUB;	
		and tandem	EPO; JPO;	
			DERWENT	
-	34	(gene ADJ silencin\$5) SAME (animal\$2 or	USPAT;	2002/09/08 13:03
		mammal\$5)	US-PGPUB;	
			EPO; JPO;	
	10	(DERWENT	0000/00/00 00 00
-	10	(gene ADJ silencin\$5).clm.	USPAT;	2002/09/08 13:03
			US-PGPUB;	
			EPO; JPO; DERWENT	
	369	gene ADJ silencin\$5	USPAT;	2002/10/31 11:44
	307	2010 1100 DITOUGHING	US-PGPUB;	2002/10/31 11:44
			EPO; JPO;	
			DERWENT	
-	2744	DOUBLE ADJ. STRANDED ADJ RNA	USPAT;	2002/11/07 11:38
			US-PGPUB;	
			EPO; JPO;	
		·	DERWENT	
-	2034	(DOUBLE ADJ STRANDED ADJ RNA) and	USPAT;	2002/10/31 11:59
		(inhibition or silen\$5)	US-PGPUB;	
			EPO; JPO;	
_	1858	((DOUBLE ADJ STRANDED ADJ RNA) and	DERWENT	2002/10/21 11.50
	1000	(inhibition or silen\$5)) and (animal or	USPAT; US-PGPUB;	2002/10/31 11:59
		mammal)	EPO; JPO;	
		31031310 ± /	DERWENT	
-	1803	(((DOUBLE ADJ STRANDED ADJ RNA) and	USPAT;	2002/10/31 12:00
		(inhibition or silen\$5)) and (animal or	US-PGPUB;	. , .,
		mammal)) and gene	EPO; JPO;	
			DERWENT	
-	1847	(((DOUBLE ADJ STRANDED ADJ RNA) and	USPAT;	2002/10/31 12:01
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		mammal)) and gene\$5) and (antisense or	EPO; JPO;	
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-	4	((((DOUBLE ADJ STRANDED ADJ RNA) and	USPAT;	2002/10/31 12:02
		(inhibition or silen\$5)) and (animal or	US-PGPUB;	
		mammal)) and gene\$5) and (antisense or	EPO; JPO;	
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-	0	WO ADJ "941550"	USPAT;	2002/11/07 11:34
			US-PGPUB;	
			EPO; JPO;	
_	6	WO ADJ "9401550"	DERWENT USPAT;	2002/11/07 11:36
	١	"O 1100 \J401330	US-PGPUB;	2002/11/0/ 11:36
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			DERWENT	
-	121	AGRAWAL-Sudhir.in.	USPAT;	2002/11/07 11:37
			US-PGPUB;	
			EPO; JPO;	
L			DERWENT	

-	119	AGRAWAL-Sudhir.in. and oligonucleotide	USPAT;	2002/11/07 11:38
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			EPO; JPO;	
			DERWENT	
-	75	(AGRAWAL-Sudhir.in. and oligonucleotide)	USPAT;	2002/11/07 11:41
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			EPO; JPO;	
	-		DERWENT	
-	12	((AGRAWAL-Sudhir.in. and oligonucleotide)	USPAT;	2002/11/07 11:46
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			EPO; JPO;	
			DERWENT	
-	1	435/325.ccls. and AGRAWAL-Sudhir.in.	USPAT;	2002/11/07 11:49
ļ			US-PGPUB;	
			EPO; JPO;	
			DERWENT	
-	2	435/320.1.ccls. and AGRAWAL-Sudhir.in.	USPAT;	2002/11/07 11:49
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	1		EPO; JPO;	
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-	75	AGRAWAL-Sudhir.in. and (sense or	USPAT;	2002/11/07 11:50
		antisense)	US-PGPUB;	
			EPO; JPO;	
			DERWENT	
-	10	AGRAWAL-Sudhir.in. and (sense or	USPAT;	2002/11/07 11:50
1		antisense).clm.	US-PGPUB;	
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			DERWENT	
-	9	AGRAWAL-Sudhir.in. and (SELF-STABILIZED	USPAT;	2002/11/07 11:56
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			EPO; JPO;	
			DERWENT	

(FILE 'HOME' ENTERED AT 10:45:14 ON 12 NOV 2002)

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FILE 'MEDLINE, AGRICOLA, CANCERLIT, SCISEARCH, CAPLUS, EMBASE, BIOSIS,
     MEDICONF' ENTERED AT 10:45:23 ON 12 NOV 2002
           40392 S GENE (L) SILEN?
L1
L2
            3154 S L1 AND (TANDEM OR REPEAT)
1.3
              75 S L2 AND (SENSE OR ANTISENSSE)
              29 DUP REM L3 (46 DUPLICATES REMOVED)
L4
L5
              29 FOCUS L4 1-
             457 S D HIS
L<sub>6</sub>
               9 S L5 AND PY<=1998
Ь7
               0 S L4 AND GRAHAM?/AU
\Gamma8
                 E GRAHAM MICHAEL?/AU
L9
              20 S E1
               5 S E2
L10
              25 S L9 OR L10
L11
              18 DUP REM L11 (7 DUPLICATES REMOVED)
L12
L13
               0 S L12 AND L4
               0 S L12 AND L2
L14
               3 S L12 AND L1
L15
=> d an ti so au ab pi 115 1-3
L15 ANSWER 1 OF 3 CAPLUS COPYRIGHT 2002 ACS
AN
     2001:713532 CAPLUS
DN
     135:268121
     Post-transcriptional gene silencing via reduction of a
     target transcript translation for manipulation in the phenotype of an
     animal
     PCT Int. Appl., 176 pp.
SO
     CODEN: PIXXD2
IN
     Graham, Michael Wayne; Rice, Robert Norman; Murphy, Kathleen
     Margaret; Reed, Kenneth Clifford
AΒ
     The present invention relates generally to a method of inducing, promoting
     or otherwise facilitating a change in the phenotype of an animal cell or
     group of animal cells including a animal comprising said cells.
     modulation of phenotypic expression is conveniently accomplished via
     genotypic manipulation through such means as reducing translation of a
     target transcript (co-suppression). One aspect of the present invention
     provides a genetic construct comprising a nucleotide sequence
     substantially identical to a target endogenous gene of a
     vertebrate animal cell, and further comprises a nucleotide sequence
     complementary to said target gene, wherein the sequences
     identical and complementary to said target gene are sepd. by an
     intron sequence. In prefered embodiment said intron sequence is an intron
     from a gene encoding .beta.-globin, and even more preferred the .beta.-globin intron is human .beta.-globin intron 2. The ability to
     induce, promote or otherwise facilitate the silencing of
     expressible genetic sequences provides a means for modulating the
     phenotype in, for example, the medical or veterinary industries.
     Expressible genetic sequences contemplated by the present invention
     including not only genes normally resident in a particular
     animal cell (i.e. indigenous genes) but also genes
     introduced through recombinant means or through infection by pathogenic
     agents such as viruses.
                   KIND DATE
     PATENT NO.
                                             APPLICATION NO. DATE
                                            WO 2001-AU297
                       A1 20010927
                                                               20010316
     WO 2001070949
         W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN,
              CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM,
              HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS,
              LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO,
         RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM
RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY,
              DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF,
              BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
L15 ANSWER 2 OF 3 CAPLUS COPYRIGHT 2002 ACS
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2001:558239 CAPLUS

- TI Suppression of **gene silencing**: a threat to virus-resistant transquenic plants
- SO Trends Plant Sci. (2001), 6(5), 246-247 CODEN: TPSCF9; ISSN: 1360-1385
- AU Mitter, Neena; Sulistyowati, Emy; Graham, Michael W.; Dietzgen, Ralf G.
- AB Unavailable
- L15 ANSWER 3 OF 3 CAPLUS COPYRIGHT 2002 ACS
- AN 1998:745287 CAPLUS
- DN 130:63748
- TI Virus resistance and **gene silencing** in plants can be induced by simultaneous expression of sense and antisense RNA
- SO Proceedings of the National Academy of Sciences of the United States of America (1998), 95(23), 13959-13964
 CODEN: PNASA6; ISSN: 0027-8424
- AU Waterhouse, Peter M.; Graham, Michael W.; Wang, Ming-Bo
- AΒ Many examples of extreme virus resistance and posttranscriptional gene silencing of endogenous or reporter genes have been described in transgenic plants contg. sense or antisense transgenes. In these cases of either cosuppression or antisense suppression, there appears to be induction of a surveillance system within the plant that specifically degrades both the transgene and target RNAs. Transforming plants with virus or reporter gene constructs that produce RNAs capable of duplex formation confer virus immunity or gene silencing on the plants. This was accomplished by using transcripts from one sense gene and one antisense gene colocated in the plant genome, a single transcript that has self-complementarity, or sense and antisense transcripts from genes brought together by crossing. A model is presented that is consisted with these data and those of other workers, describing the processes of induction and execution of posttranscriptional gene silencing.

(FILE 'HOME' ENTERED AT 10:45:14 ON 12 NOV 2002)

FILE 'MEDLINE, AGRICOLA, CANCERLIT, SCISEARCH, CAPLUS, EMBASE, BIOSIS, MEDICONF' ENTERED AT 10:45:23 ON 12 NOV 2002 40392 S GENE (L) SILEN? L1 L23154 S L1 AND (TANDEM OR REPEAT) L3 75 S L2 AND (SENSE OR ANTISENSSE) 29 DUP REM L3 (46 DUPLICATES REMOVED) L4L5 29 FOCUS L4 1-L6 457 S D HIS 9 S L5 AND PY<=1998 1.7 => d an ti so au ab 17 2 3 ANSWER 2 OF 9 AGRICOLA 1.7 1999:22365 AGRICOLA AN A transgene with repeated DNA causes high frequency, post-transcriptional ΤI suppression of ACC-oxidase gene expression in tomato. The Plant journal : for cell and molecular biology, Sept 1998. Vol. 15, No. 6. p. 737-746 Publisher: Oxford: Blackwell Sciences Ltd. ISSN: 0960-7412 Hamilton, A.J.; Brown, S.; Yuanhai, H.; Ishizuka, M.; Lowe, A.; Solis, A.G.A.; Grierson, D. Gene silencing with sense genes is an important method for down-regulating the expression of endogenous plant genes, but the frequency of silencing is unpredictable. Fifteen per cent of tomato plants transformed with a 35S-ACC-oxidase (ACO1) sense gene had reduced ACC-oxidase activity. However, 96% of plants transformed with an ACC-oxidase sense gene, containing two additional upstream inverted copies of its 5' untranslated region, exhibited reduced ACC-oxidase activity compared to wild-type plants. In the three plants chosen for analysis, there were substantially reduced amounts of both endogenous and transgenic ACO RNA, indicating that this was an example of co-suppression. Ribonuclease protection assays using probes spanning intron-exon borders showed that the reduced accumulation of endogenous ACO mRNA occurred post-transcriptionally since the abundance of unprocessed transcripts was not affected. The ACO1 transgene with the repeated 5'UTR also strongly inhibited the accumulation of RNA from the related ACO2 gene in flowers, although there is little homology between the 5'UTRs of ACO1 and ACO2. These results indicate that although repeated DNA in a transgene greatly enhances the probability of gene silencing of an endogenous gene, it also involves generation of a trans-acting silencing signal produced, at least partly, from sequences external to the repeat. L7 ANSWER 3 OF 9 AGRICOLA 1998:28715 AGRICOLA AN TΙ Post-transcriptional silencing of chalcone synthase in Petunia by inverted transgene repeats. so The Plant journal : for cell and molecular biology, July 1997. Vol. 12, No. 1. p. 63-82 Publisher: Oxford : Blackwell Sciences Ltd. ISSN: 0960-7412 ΔII Stam, M.; Bruin, R. de.; Kenter, S.; Hoorn, R.A.L. van der.; Blockland, R. van.; Mol, J.N.M.; Kooter, J.M. AB To induce post-transcriptional silencing of flower pigmentation genes by homologous sense transgenes in transgenic petunias, it is not necessary for the transgenes to be highly transcribed. Even promoterless transgenes can induce silencing. Here it is shown that in these cases silencing is mediated by multimeric transgene/T-DNA loci in which the T-DNAs are arranged as inverted repeats (IRs). With the transgene constructs used, monomeric T-DNA loci are unable to confer silencing even though they modulate IR-induced silencing. IRs with the silencing sequences

proximal to the centre (IRc) induce a more severe silencing than

silencing, as observed in a side branch of one of the chalcone

IRs with these sequences distal to the centre (IRn). Somatic reversion of

synthase (Chs) transformants, was associated with a deletion of the IR

locus from L1 cells, the meristematic cell layer that expresses the endogenous Chs genes in the flower corolla. Taken together, these data indicate that the post-transcriptional silencing mechanism can be activated by inverted transgene repeats. It is also shown that a silent IR UidA-ChsA locus silences the expression of a monomeric 35S promoter-driven UidA-ChsA transgene only in corollas where the endogenous Chs genes are highly transcribed. These results are consistent with a model in which an IR, by virtue of its palindromic sequence organization, is able to promote the production of aberrant RNAs from the endogenous homologs as a result of ectopic pairing.

ANSWER 1 OF 29 CAPLUS COPYRIGHT 2002 ACS 2002:142846 CAPLUS AN DN 136:178951 Improved methods of gene silencing in plant using inverted repeat sequences from NOS gene SO PCT Int. Appl., 39 pp. CODEN: PIXXD2 IN Gutterson, Neal; Oeller, Paul The present invention provides methods for inhibiting target gene AB expression, by expressing in a cell a nucleic acid construct comprising an inverted repeat and a sense or antisense region having substantial sequence identity to a target gene, wherein the inverted repeat is unrelated to the target gene. The inverted repeat is chosen from any suitable sequence and has the ability to form a double stranded RNA in the cell. N another preferred embodiment, the heterologous inverted repeat of the invention is from Agrobacteriumn tumefaciens NOS gene or from the 3' untranslated region of the NOS gene. In one embodiment, the improved gene silencing construct is expressed in a plant cell, where the transcript, or fragments thereof, is taken up by plant pathogens such as fungi, bacteria, nematodes, e.g., cyst and root knot nematodes, and insects, e.g., sucking insects, leading to gene silencing in the pathogen. In another embodiment, the improved gene silencing construct is expressed in a transgenic plant, and is used to regulate expression of the transgene. N another embodiment, the gene silencing vector is used to regulate expression of an endogenous plant gene, e.g., to regulate plant phenotypes such as disease resistance. PATENT NO. KIND DATE APPLICATION NO. DATE PТ WO 2002014472 A2 20020221 WO 2001-US25538 20010814 WO 2002014472 Α3 20020718 W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, $\mathtt{UZ},\ \mathtt{VN},\ \mathtt{YU},\ \mathtt{ZA},\ \mathtt{ZW},\ \mathtt{AM},\ \mathtt{AZ},\ \mathtt{BY},\ \mathtt{KG},\ \mathtt{KZ},\ \mathtt{MD},\ \mathtt{RU},\ \mathtt{TJ},\ \mathtt{TM}$ RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG A5 AU 2001088257 20020225 AU 2001-88257 20010814 L5 ANSWER 2 OF 29 CAPLUS COPYRIGHT 2002 ACS AN 2000:718761 CAPLUS DN134:203186 ΤI Gene expression: Total silencing by intron-spliced SO Nature (London) (2000), 407(6802), 319-320 CODEN: NATUAS; ISSN: 0028-0836 ΑU Smith, Neil A.; Singh, Surinder P.; Wang, Ming-Bo; Stoutjesdijk, Peter A.; Green, Allan G.; Waterhouse, Peter M. Post-transcriptional gene silencing (PTGS), a sequence-specific RNA degrdn. mechanism inherent in many life forms, can be induced in plants by transforming them with either antisense or co-suppression constructs, but typically this results in only a small proportion of silenced individuals. Here we show that gene constructs encoding intron-spliced RNA with a hairpin structure can induce PTGS with almost 100% efficiency when directed against viruses or endogenous genes. Using principles we developed for silencing constructs that express double-stranded RNA and inverted-repeat RNA, we made a construct encoding a single self complementary hairpin RNA (hpRNA) of the Niaprotease (Pro) gene sequence of potato virus Y (PVY). The construct contains sense and antisense Pro sequences flanking a nucleotide spacer fragment derived from uidaA (GUS) gene. About 60% of the plants that are transformed with the constructs were immune to the virus. In the next expt., we replaced the spacer with an intron sequence, which is spliced out during pre-mRNA processing to produce loopless hpRNA. As a

SK-1636

control, the intron sequence was inserted in the reverse, non-splicing, orientation. When transformed into tobacco, 22 of 34 (65%) reverse-intron plants were immune, a similar frequency to plants transformed with the GUS spacer construct. Amazingly, 22 of 23 plants transformed with the construct contg. the functional intron were immune to the virus. This same enhancement was obsd. when hpRNA constructs against the endogenous approch.12-desaturase (Fad2) gene of Arabidopsis, in which 100% (30/30) of plants transformed with the intron construct showed silencing of the gene. The process of intron excision from the construct by spliceosome might help to align the complementary arms to the hairpin in an environment favoring RNA hybridization, promoting the formation of a duplex. Alternatively, splicing may transiently increase the amt. of hairpin RNA by facilitating, or retarding, the hairpin's passage from the nucleus, or by creating a smaller, less nuclease-sensitive loop.

L5 ANSWER 3 OF 29 CAPLUS COPYRIGHT 2002 ACS

AN 2001:507844 CAPLUS

DN 135:88019

TI Compositions and methods for **gene silencing** by expression of double-stranded RNA

SO PCT Int. Appl., 67 pp. CODEN: PIXXD2

IN Driggell Moniga, Taxorna

IN Driscoll, Monica; Tavernarakis, Nektarios

AΒ DNA constructs are provided for disrupting gene expression in targeted organisms, including humans, mice, plants, insects, and nematodes. The DNA constructs involve a transcription promoter followed by a gene coding sequence in the sense orientation linked to the same coding sequence in an antisense orientation followed by a transcription terminator. Use of a DNA construct, which is an inverted repeat (IR) of a gene cloned in an expression vector, for treatment of Alzheimer's and Parkinson's disease and tomato leaf curl virus is claimed. RNA interference by double-stranded RNA using methods claimed in this invention was demonstrated in Caenorhabditis elegans and was more effective compared to gene disruption methods such as injection of dsRNA and expression of an antisense DNA strand alone. For some but not all genes tested, transgenic C. elegans lines contg. extrachromosomal IR gene constructs under control of the heat shock-inducible promoter hsp16-2 had high percentages of progeny with the predicted phenotype for deletions of the gene used in the construct. C. elegans gene C37A2.5 required for development past the L2 larval stage, gene F26F12.7 required for fertility, and gene mec-4 required for touch sensitivity could be disrupted by the claimed methods while genes efk-1 and unc-119 were not affected. The ability of an hsp16-2 promoter-green fluorescent protein (GFP) gene IR construct to affect expression of an integrated GFP gene in C. elegans was also demonstrated.

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PATENT NO. KIND DATE APPLICATION NO. DATE

WO 2001049844 A1 20010712 WO 2001-US126 20010102

W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM

RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
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- L5 ANSWER 4 OF 29 CAPLUS COPYRIGHT 2002 ACS
- AN 2002:575187 CAPLUS
- DN 137:122370
- TI Sense and antisense constructs for silencing of barley yellow dwarf virus-PAV RNA dependant RNA polymerase and improved viral resistance of cereal plants
- SO PCT Int. Appl., 56 pp. CODEN: PIXXD2
- IN Waterhouse, Peter; Wang, Ming-Bo; Abbott, David
- AB The invention provides a DNA mol. comprising a plant-operable promoter operably linked to a DNA region that is capable of being transcribed in

SK-1636

the cells of a cereal plant to produce RNA comprising inverted repeat sequence least about 19 nucleotides from the sequence of an RNA dependent RNA polymerase gene of Barley Yellow Dwarf Virus (BYDV). These constructs encode sense and antisense RNA mols. and are directed to the RNA dependent RNA polymerase gene. The RNA is capable of forming an artificial hairpin RNA structure with a double stranded RNA stem by base-pairing between the regions with sense and antisense nucleotide sequence such that at least the 19 consecutive nucleotides of the sense sequence base pair with the 19 consecutive nucleotides of the antisense sequence resulting in an artificial hairpin structure. The DNA mol. is useful for reducing expression of the viral RNA dependant RNA polymerase and for enhancing the resistance of cereal crops to BYDV, optionally in the presence of a viral product that normally inactivates other modes of post transcriptional gene silencing. Methods are provided for producing transgenic cereal plant lines comprising the DNA mol. of the invention integrated into their genome, and selecting those lines having resistance to BYDV infection. Transgenic plants having enhanced resistance to BYDV are also provided.

PATENT NO. KIND DATE APPLICATION NO. DATE

PI WO 2002059257 A2 20020801 WO 2001-IB2737 20011031

W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG

- L5 ANSWER 5 OF 29 MEDLINE
- AN 2001418932 MEDLINE
- Double-stranded RNA-mediated silencing of genomic tandem repeats and transposable elements in the D. melanogaster germline. SO CURRENT BIOLOGY, (2001 Jul 10) 11 (13) 1017-27.
- Journal code: 9107782. ISSN: 0960-9822. AII Aravin A A; Naumova N M; Tulin A V; Vagin V V; Rozovsky Y M; Gvozdev V A BACKGROUND: The injection of double-stranded RNA (dsRNA) has been shown to induce a potent sequence-specific inhibition of gene function in diverse invertebrate and vertebrate species. The homology-dependent posttranscriptional gene silencing (PTGS) caused by the introduction of transgenes in plants may be mediated by dsRNA. The analysis of Caenorhabditis elegans mutants impaired with dsRNA-mediated silencing and studies in plants implicate a biological role of dsRNA-mediated silencing as a transposon-repression and antiviral mechanism. RESULTS: We investigated the silencing of testis-expressed Stellate genes by paralogous Su(Ste) tandem repeats, which are known to be involved in the maintenance of male fertility in Drosophila melanogaster. We found that both strands of repressor Su(Ste) repeats are transcribed, producing sense and antisense RNA. The Stellate silencing is associated with the presence of short Su(Ste) RNAs. Cotransfection experiments revealed that Su(Ste) dsRNA can target and eliminate Stellate transcripts in Drosophila cell culture. The short fragment of Stellate gene that is homologous to Su(Ste) was shown to be sufficient to confer Su(Ste)-dependent silencing of a reporter construct in testes. We demonstrated that Su(Ste) dsRNA-mediated silencing affects not only Stellate expression but also the level of sense Su(Ste) RNA providing a negative autogenous regulation of Su(Ste) expression. Mutation in the spindle-E gene relieving Stellate silencing also leads to a derepression of the other genomic tandem repeats and retrotransposons in the germline. CONCLUSIONS: Homology-dependent gene silencing was shown to be used to inhibit Stellate gene expression in the D. melanogaster germline, ensuring male

fertility. dsRNA-mediated **silencing** may provide a basis for negative autogenous control of **gene** expression. The related surveillance system is implicated to control expression of